Dickkopf-1: As a Diagnostic and Prognostic Serum Marker for Hepatocellular Carcinoma

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Background and study aim: Hepatocellular carcinoma (HCC) accounts for 70 - 80% of all liver cancers and the 5-year survival is only 3 - 5%. This bad prognosis is due to the lack of an effective method for early diagnosis. So, only 30 - 40% of patients with HCC are suitable for curative treatments at the time of diagnosis. Thus, there is a great need for tools to diagnose HCC early especially in cirrhotic patients. The aim of this work is to assess the validity of serum DKK1 as a diagnostic marker for HCC and to assess prognostic value of serum DKK1 in predicting treatment response, complication and survival in HCC patients.

Patients and Methods: This study included 60 Patients divided into two groups. Group A: consisted of 30 patients with post hepatitis C and/or B liver cirrhosis. Group B: consisted of 30 patients with HCC on top of post hepatitis C and/or B liver cirrhosis. Group B patients underwent either radiofrequency ablation or ethanol injection. Clinical assessment, routine laboratory evaluation, CT studies and measurement of serum alpha-fetoprotein (AFP) and DKK1 were performed to all patients and repeated to group B patients 1 and 3 months after treatment.

Results: The optimum cut off value of DKK1 for diagnosis of HCC was 4.3 ng/mL (AUC 0.89, sensitivity 66.7% and specificity 96.6%) (P<0.001). While, the optimum cut off value for AFP was > 101 ng/mL with 90% sensitivity and 75.9% specificity (p<0.001). Testing of both DKK1 and AFP increased the diagnostic accuracy for HCC (AUC 0.901, sensitivity 93.3%, and specificity 75.9) (P<0.001). Serum DKK1 level significantly decreases after HCC treatment with either radiofrequency ablation or ethanol injection (P<0.001).

Conclusion: Testing of both DKK1 and AFP significantly increased the diagnostic accuracy for HCC. Meanwhile, DKK1 can be used alone for HCC diagnosis even in HCC with inconclusive AFP. DKK1 has a promising prognostic value and can be used for follow up of HCC patients who underwent loco-regional treatment.

INTRODUCTION

Hepatocellular carcinoma (HCC) is the sixth most common malignant disease and the third leading cause of cancer-related death worldwide. HCC is prevalent in Asia and Africa, but recently it raises in the Western world due to an increase in hepatitis C virus (HCV) infection [1]. In Egypt, Liver cancer forms 11.75% of the malignancies of all digestive organs and 1.6% of total malignancies [2,3]. Risk factors for HCC include chronic hepatitis B virus (HBV) and chronic hepatitis C infections, cirrhosis, chronic alcohol abuse, aflatoxin ingestion, non-alcoholic steatohepatitis and metabolic liver diseases [4]. Both HCV and HBV infections are the most common risk factors for HCC among Egyptian patients. 10%-20% of the general Egyptian populations are infected with HCV [5]. 80% - 90% of HCC patients have underlying cirrhosis and the remaining 10% - 20% of cases develop HCC without cirrhosis [6,7].
HCC is a disease with fast infiltrating growth and poor prognosis [8]. The commonly used screening methods for liver cancer are ultrasound examination of the liver and determination of serum AFP level [9]. Abdominal ultrasound is a better, simple and easy method for detection of HCC but it is operator dependent and many focal lesions can be missed [10]. AFP has approximately 60% specificity and 40% sensitivity for HCC diagnosis, since minor elevations are common in patients with chronic liver disease, cirrhosis, germ cell tumors and in pregnancy [11]. So, it is necessary to find a specific & sensitive marker for early diagnosis of HCC and for monitoring of treatment response.

Dickkopf-1 (DKK1) is a secretory protein which was identified in 1998. DKK1 is an inhibitor of Wnt/β-catenin signalling and a downstream target of β-catenin [12]. The Wnt/β-catenin signalling pathway plays main role in development of both normal liver and hepatic carcinogenesis [13]. It is hardly expressed in normal human adult tissues except in placental and embryonic tissues [14]. DKK1 is up regulated in various cancers including breast, lung, ovarian, prostate cancers and HCC [15].

This work aimed to assess validity of serum DKK1 as a diagnostic marker for HCC and to assess prognostic value of serum DKK1 in predicting treatment response, complication and survival in HCC patients.

PATIENTS AND METHODS

This case control study was conducted in Tropical Medicine and Clinical Pathology Departments, Faculty of Medicine, Zagazig University Hospitals, Egypt during the period from January 2014 till March 2016.

This study included 60 Patients divided into two groups: Group A: consisted of 30 patients with post hepatitis C and/or B liver cirrhosis. Group B: consisted of 30 patients with HCC on top of post hepatitis C and/or B liver cirrhosis.

Inclusion criteria

Group (A) included cirrhotic patients with no evidence of hepatic focal masses in ultrasound evaluation. Cirrhotic patients are child class A or B according to Child Pugh score. Patients with liver cirrhosis were diagnosed by liver biopsy, laboratory and/or imaging evidence including (nodular liver contour, presence of ascites, portal hypertension, varices, enlargement of the caudate lobe, splenomegaly and collateral portal venous anastomoses).

Group (B) included patients with HCC on top of cirrhotic liver. HCC was diagnosed by CT criteria (filling of the dye in arterial phase and rapid fade out in venous and delayed phases) and/or by histopathology according to the American Association for the Study of Liver Diseases guidelines. HCC patients will be Child class A or B according to Child Pugh score for cirrhotic patients.

Exclusion criteria

Patients who had any other tumors or history of other tumors were excluded from the study. Also, patients with Child-Pugh class C, vascular invasion or extra hepatic metastasis were excluded from the study.

All patients were subjected to full history, complete physical examination and laboratory investigation in the form of liver function tests, kidney function tests, complete blood count, AFP, viral markers (HBs Ag and HCV Abs) and serum DKK1. Also, all patients were subjected to abdominal ultrasound. HCC was diagnosed by triphasic CT examination of the abdomen or by liver biopsy (FNAB) (imaging is not conclusive). Group B patients underwent either radio-frequency ablation or ethanol injection according to the Barcelona Clinic Liver Cancer (BCLC) staging system and followed up by laboratory investigations (CBC, LFTs, KFTs, AFP, and DKK1), abdominal ultrasound and triphasic CT scan 1 and 3 months after treatment.

Dickkopf-1 (DKK1)

It was determined by Human Dickkopf-1(DKK1) ELISA Kits provided by WKEA MED SUPPLIES CORP, USA, according to the manufacturer’s protocol. This kit allows for the determination of DKK1 concentrations in Human serum, plasma, and other biological fluids.

The kit assay Human DKK1 level in the sample, by using Purified Human DKK1 antibody to coat microtiter plate wells, make solid-phase antibody, then add DKK1 to wells, Combined DKK1 antibody which With enzyme labeled, become antibody - antigen - enzyme-antibody complex, after washing Completely, Add substrate, substrate becomes blue color At HRP enzyme-catalyzed, reaction is terminated by the addition of a sulphuric acid solution and the color change is...
measured spectrophotometrically at a wave length of 450 nm.

The concentration of DKK1 in the samples is then determined by comparing the O.D. of the samples to the standard curve.

**Ethanol injection**

All lesions were injected by absolute alcohol; ultrasound guided in multiple sessions, once weekly, under complete aseptic condition and 10 mg midazolam as a sedative agent.

The same operator used spinal needle (20 gauges) to inject ethanol intra-lesionally and leave the needle for 2 minutes in place, then injection of local anesthetic during withdrawal of the needle to minimize the irritant effect of refluxed ethanol to the capsule.

The total amount of ethanol can be calculated according to the following equation:

\[
V = \frac{4}{3}\pi (r + 0.5)^3
\]

Where: \(V\) = Volume of ethanol, \(\pi = \frac{22}{7}\), \(r\) = radius of the tumor by cm plus 0.5 cm as safety margin. The average amount per session was 6.8 cc, with average 5 sessions per lesion and average amount of 35 cc per lesion [16].

**Radiofrequency ablation**

All patients were fasted before the procedure. Treatment was performed with sedation using midazolam (Dormicum R 10 mg amp; Roche) 0.03-0.1 mg/kg/IV every 30 minutes, propofol (Diprivan R 20 mg amp; Astra) 0.5 mg/kg/IV over 3-5 minutes.

All lesions were ablated by the same operator hands, under complete aseptic condition at Ultrasonography Unit, Tropical medicine department. Multiple curved, retractable electrodes are kept inside the needle until its tip is positioned within a tumor. When properly positioned, a plunger on the hub of the needle is advanced so that the electrodes extend from the needle tip. Multiple electrode tips of an expanding electrode are active. This results in more homogenous heat distribution within the tumor and creates a reproducible sphere of ablation every time. Patients were observed for 6 hours for blood pressure, pulse, pain and vomiting.

**Statistical analysis**

All data were collected, tabulated and statistically analyzed using SPSS 20.0 for windows. Quantitative data were expressed as the mean ± SD & median (range), and qualitative data were expressed as an absolute frequencies “number”& relative frequencies (percentage). Independent samples Student’s t-test, Mann-Whitney U, Paired t-test and Wilcoxon signed ranks test were used when needed. Percent of categorical variables were compared using the Pearson’s Chi-square test or Fisher’s exact test when was appropriate. Receiver operating characteristic (ROC) curve analysis was used to identify optimal cut-off values of AFP and DKK1 with maximum sensitivity and specificity for diagnosis HCC and prediction of response. Area Under Curve (AUC) was also calculated. \(P<0.05\) was considered statistically significant (S).

**RESULTS**

This study showed no statistically significant difference between groups A & B as regard age, sex, viral etiology and Child Pugh score. Most of our patients were males (42 patients) and HCV positive (51). Table (1) showed no statistically significant difference between group A and group B as regard laboratory data except for platelet count, DKK1 and AFP (138.93 ± 37.17 Vs 110.76 ± 38.61 \(P = 0.006\)), (2.28 ± 0.90 ng/ml Vs 4.97 ± 2.23 ng/ml \(P<0.001\)) and (70.38 ± 80.52 ng/ml Vs 361.93 ± 289.91 \(P<0.001\)), respectively. We found that serum DKK1 was more elevated in HCC patients with focal lesions >3 cm than focal lesions <3 cm (6.09 ± 1.77 Vs 2.75 ± 1.09) \(P<0.001\) (Table 2).

The optimum diagnostic cut off value for DKK1 was >4.3 ng/mL with 66.7% sensitivity and 96.6% specificity while, the cut off value of AFP was >101 ng/mL with 90% sensitivity and 75% specificity for HCC diagnosis vs. cirrhotic patients \((P<0.001)\). Testing of both DKK1 and AFP increased the diagnostic accuracy for HCC compared with either test alone \((AUC 0.901, 95\% CI 0.795-0.964,\text{ sensitivity 93.3%}, \text{ and specificity 75.9}) (P<0.001)\) (Table 3; Fig. 3).

Table (4) showed no statistically significant difference among studied group as regard bilirubin, albumin, PT, creatinine and CBC before and after treatment, whereas DKK1, AFP, ALT and AST showed statistically significant improvement in these patients after treatment. DKK1 levels before and after treatment were 4.97 ± 2.23 ng/ml and 2.75 ± 1.52 ng/ml respectively \((p<0.001)\). This study showed highly statistically significant decline of DKK1 level among complete responder's patients \((P<0.001)\) (Table 5). The cut off value of DKK1 (before treatment) for prediction of complete response to treatment was \(\leq 5.67\) ng/mL \((p<0.001)\) (Table 6; Fig. 4).
Table (1): Laboratory investigations and tumor markers of both groups

<table>
<thead>
<tr>
<th>Laboratory findings</th>
<th>Group A (N=30)</th>
<th>Group B (N=30)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>AST (U/L)</td>
<td>64.41 ± 18.29</td>
<td>70.46 ± 31.65</td>
<td>0.756 (NS)</td>
</tr>
<tr>
<td>ALT (U/L)</td>
<td>64.06 ± 18.54</td>
<td>60.60 ± 24.98</td>
<td>0.391 (NS)</td>
</tr>
<tr>
<td>Bilirubin (mg/dl)</td>
<td>1.70 ± 0.76</td>
<td>1.93 ± 0.77</td>
<td>0.140 (NS)</td>
</tr>
<tr>
<td>Albumin (g/dl)</td>
<td>3.29 ± 0.49</td>
<td>3.29 ± 0.54</td>
<td>0.769 (NS)</td>
</tr>
<tr>
<td>PT (sec)</td>
<td>15.27 ± 1.62</td>
<td>15.83 ± 2.98</td>
<td>0.249 (NS)</td>
</tr>
<tr>
<td>Creatinine (mg/dl)</td>
<td>0.93 ± 0.18</td>
<td>1.02 ± 0.27</td>
<td>0.154 (NS)</td>
</tr>
<tr>
<td>Hemoglobin (g/dl)</td>
<td>11.46 ± 0.60</td>
<td>11.83 ± 1.55</td>
<td>0.306 (NS)</td>
</tr>
<tr>
<td>Plt (x10^3/mm^3)</td>
<td>138.93 ± 37.17</td>
<td>110.76 ± 38.61</td>
<td>0.006 (S)</td>
</tr>
<tr>
<td>WBCs (x10^3/mm^3)</td>
<td>6 ± 1.65</td>
<td>5.81 ± 2.17</td>
<td>0.444 (NS)</td>
</tr>
<tr>
<td>AFP (ng/dl)</td>
<td>70.38 ± 80.52</td>
<td>361.93 ± 289.91</td>
<td>&lt;0.001 (HS)</td>
</tr>
<tr>
<td>DKK 1 (ng/dl)</td>
<td>2.28 ± 0.90</td>
<td>4.97 ± 2.23</td>
<td>&lt;0.001 (HS)</td>
</tr>
</tbody>
</table>

Table (2): Patients with focal lesion < 3cm and patients with focal lesion 3-5 cm in group (B) as regard tumor markers

<table>
<thead>
<tr>
<th></th>
<th>Group B (N=30)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>&lt;3 cm (N=10)</td>
</tr>
<tr>
<td>AFP (ng/dl)</td>
<td>422.30 ± 349.55</td>
</tr>
<tr>
<td>DKK 1 (ng/dl)</td>
<td>2.75 ± 1.09</td>
</tr>
</tbody>
</table>

Fig. (1): Percentage of increased level of AFP and DKK 1 among HCC patients (group B)
Fig. (2): Percentage of increased level of DKK1 among HCC patients with non-conclusive AFP

Table (3): Validity of DKK1, AFP and DKK1+AFP as diagnostic markers for HCC vs. cirrhotic patient without HCC

<table>
<thead>
<tr>
<th>Cut-off value</th>
<th>Sens. % (95% CI)</th>
<th>Spec. % (95% CI)</th>
<th>PPV % (95% CI)</th>
<th>NPV % (95% CI)</th>
<th>AUC (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>DKK1 &gt;4.3 ng/mL</td>
<td>66.7% (47.2—82.7)</td>
<td>96.6% (82.2-99.9)</td>
<td>95.2% (76.2-99.9)</td>
<td>73.7% (56.9-86.6)</td>
<td>0.895</td>
</tr>
<tr>
<td>AFP &gt;101 ng/mL</td>
<td>90% (73.5-97.9)</td>
<td>75.9% (56.5-89.7)</td>
<td>79.4% (62.1-91.3)</td>
<td>88% (968.8-97.5)</td>
<td>0.895</td>
</tr>
<tr>
<td>DKK1+AFP &gt;102.2ng/mL</td>
<td>93.3% (77.9-99.2)</td>
<td>75.9% (56.5-89.7)</td>
<td>80% (63.1-91.6)</td>
<td>91.7% (73-99)</td>
<td>0.901</td>
</tr>
</tbody>
</table>

Fig. (3): ROC curve of DKK1, AFP and DKK1+AFP as diagnostic markers for HCC vs. cirrhotic patients without HCC
Table (4): Laboratory findings before and after treatment in Group B

<table>
<thead>
<tr>
<th>Laboratory findings</th>
<th>Before treatment (N=30)</th>
<th>After treatment (N=30)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>AST (U/L)</td>
<td>70.46 ± 31.65</td>
<td>55.26 ± 25.12</td>
<td>0.003(S)</td>
</tr>
<tr>
<td>ALT (U/L)</td>
<td>60.60 ± 24.98</td>
<td>55.86 ± 26.17</td>
<td>0.004(S)</td>
</tr>
<tr>
<td>Bilirubin (mg/dl)</td>
<td>1.93 ± 0.77</td>
<td>2.05 ± 0.74</td>
<td>0.131(NS)</td>
</tr>
<tr>
<td>Albumin (g/dl)</td>
<td>3.29 ± 0.54</td>
<td>3.08 ± 0.45</td>
<td>0.129(NS)</td>
</tr>
<tr>
<td>PT (sec)</td>
<td>15.83 ± 2.98</td>
<td>17.20 ± 3.99</td>
<td>0.057(NS)</td>
</tr>
<tr>
<td>Creatinine (mg/dl)</td>
<td>1.02 ± 0.27</td>
<td>1.07 ± 0.24</td>
<td>0.07(NS)</td>
</tr>
<tr>
<td>Hemoglobin (g/dl)</td>
<td>11.83 ± 1.55</td>
<td>11.23 ± 1.45</td>
<td>0.16(NS)</td>
</tr>
<tr>
<td>Plt (x10^3/mm^3)</td>
<td>110.76 ± 38.61</td>
<td>109.5 ± 36.62</td>
<td>0.231(NS)</td>
</tr>
<tr>
<td>WBCs(x10^3/mm^3)</td>
<td>5.81 ± 2.17</td>
<td>5.10 ± 1.99</td>
<td>0.18(NS)</td>
</tr>
<tr>
<td>AFP (ng/dl)</td>
<td>361.93 ± 289.91</td>
<td>286.93 ± 241.30</td>
<td>&lt;0.001(HS)</td>
</tr>
<tr>
<td>DKK 1 (ng/dl)</td>
<td>4.97 ± 2.23</td>
<td>2.75 ± 1.52</td>
<td>&lt;0.001(HS)</td>
</tr>
</tbody>
</table>

Table (5): Tumor markers 1 month after treatment among partial and complete responder's patients

<table>
<thead>
<tr>
<th>Tumor markers</th>
<th>Partial responder (N=9)</th>
<th>Complete responder (N=21)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>AFP before</td>
<td>402.11 ± 264.37</td>
<td>344.71 ± 304.75</td>
<td>0.402(NS)</td>
</tr>
<tr>
<td>AFP after</td>
<td>253.22 ± 135.39</td>
<td>301.38 ± 276.34</td>
<td>0.751(NS)</td>
</tr>
<tr>
<td>DKK 1 before</td>
<td>7.52 ± 1.44</td>
<td>3.88 ± 1.49</td>
<td>&lt;0.001(HS)</td>
</tr>
<tr>
<td>DKK 1 after</td>
<td>3.36 ± 1.57</td>
<td>2.49 ± 1.45</td>
<td>0.167(NS)</td>
</tr>
</tbody>
</table>

Table (6): Validity of DKK1 (before treatment) in prediction of complete response to treatment; ROC curve Analysis

<table>
<thead>
<tr>
<th>Cut-off value</th>
<th>Sens. % (95% CI)</th>
<th>Spec. % (95% CI)</th>
<th>PPV % (95% CI)</th>
<th>NPV % (95% CI)</th>
<th>AUC (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>DKK1 before ≤ 5.67 ng/mL</td>
<td>100% (83.9-100)</td>
<td>100% (66.4-100)</td>
<td>100% (83.9-100)</td>
<td>100% (66.4-100)</td>
<td>100% (78.7-100)</td>
</tr>
</tbody>
</table>

Fig. (4): ROC curve of DKK1 (before treatment) in prediction of complete response to treatment
DISCUSSION

HCC is usually asymptomatic in early stages and tends to be invasive. Most HCC patients are presented with non-operable disease and this makes its early diagnosis critical for a good prognosis. Early HCC detection gives the opportunity to employ curative treatments such as liver transplantation, resection or local ablative therapy, which are the best way to prolong survival [17]. So, continuous researches are ongoing worldwide to find and evaluate an early sensitive and specific marker for HCC diagnosis [18].

Protein markers that measured in serum are the most applicable tests for clinical assessments and population studies [19,20]. DKK1 is a secretory protein, specifically over expressed in cancer cells and is hardly detectable in human adult normal tissues except in placenta and embryonic tissues. Therefore, this protein might have potential as a cancer-specific serum biomarker for various human cancers including HCC [21].

In the present study, there was a statistically significant difference between the mean value of DKK1 in patients with HCC compared to patients with liver cirrhosis with mean values of 4.97±2.23 ng/mL and 2.28±0.90 ng/mL respectively. These results were in agreement with those of Shen et al., 2012 and Zhang et al., 2014 who showed that serum DKK1 level was higher in patients with HCC than cirrhotic patients, chronic hepatitis B and healthy control [22,23].

In our study, serum DKK1 level was more elevated in Child B cirrhotic patients than Child A patients (in group A) with mean level 9.95 ± 1.04 and 1.87±0.49 respectively. So, DKK1 levels increase with hepatic dysfunction. Also, serum DKK1 was more elevated in HCC patients with focal lesions >3 cm than focal lesions <3 cm (6.09±1.77 and 2.75±1.09 respectively) in group B. This indicated that DKK1 level increase with disease progression from cirrhosis to small focal lesion then large focal mass. These results agreed with those of Tung et al., (2011) who reported a stepwise increase in serum DKK1 from cirrhosis group to early HCC then to advanced HCC group [24].

In this study, ROC curves revealed that the optimum diagnostic cut off value of DKK1 is 4.3 ng/mL for diagnosis of HCC (AUC 0.895, 95% CI 0.787-0.960, sensitivity 66.7%, specificity 96.6%). This result is in agreement with that of Shen et al., (2012) and Zhang et al., (2014) who reported AUC (0.848 & 0.84), sensitivity (69.1% & 65%), specificity (90.6% & 94%) for HCC diagnosis versus cirrhosis control [22,23]. In contrast, Yang et al., (2013) showed that the DKK1 AUC (0.717) for HCC diagnosis was lower than the AUC in our study (0.895) [26].

In the present study, diagnostic cut off value for AFP was > 101 ng/mL for HCC in cirrhotic patients with 90% sensitivity, 75.9% specificity and 0.895 AUC. Serum DKK1 had similar AUC as AFP, higher specificity and lower sensitivity than AFP and this could be due to small sample size and only cirrhotic patients included as a control group not healthy control. This was in agreement with Nakamura et al., 2006 who showed that the cut off value of AFP for HCC diagnosis was 100 ng/ml with (33%) sensitivity and (99%) specificity [27]. In contrast, Farinati et al., 2006; and Debruyne and Delanghe, 2008 reported other sensitivity, specificity and cut off value for AFP for HCC diagnosis [28,29].

A greater proportion of HCC patients in our study were positive for DKK1 than for AFP. Furthermore, 8 of 13 AFP negative HCC patients had positive DKK1 result and all AFP-positive patients had +ve DKK1 results (Fig. 1; Fig. 2). The ROC curves for DKK1 indicated the diagnosis of HCC irrespective of AFP status. This finding was in agreement with that of Shen et al., (2012) and Yang et al., (2013) [22,26].

In this study, testing of both DKK1 and AFP increased the diagnostic accuracy for HCC compared with either test alone (AUC 0.901, 95% CI, 0.795-0.964, sensitivity 93.3%, and specificity 75.9).This was in agreement with Ge et al., 2015 who showed that testing of both AFP and DKK1 had AUC 0.93, sensitivity 88.8%, and specificity 88.12% [30]. In contrast, Eun et al., 2016 reported that testing of AFP and DKK1 had AUC 0.76, sensitivity 78 %, and specificity 73% [31].

Group B patients underwent either radiofrequency ablation (12 patients) or ethanol injection (18 patients) according to the Barcelona Clinic Liver Cancer (BCLC) staging system [32]. Percutaneous ablation is the preferred treatment option for patients in this study. Both radiofrequency ablation and percutaneous injection therapy have a well-documented loco-regional antitumor effect and are the most two commonly employed methods for HCC treatment [33,34]. Liver transplantation and surgical resection are the standard treatment modality to achieve a long-term survival.
However, both of them are major surgery with many complications and have negative impact on patients’ especially cirrhotic [35,36].

This study showed no statistically significant difference between patients treated with either radiofrequency ablation or ethanol injection as regards AFP and DKK1. There was decrease in mean level of both markers after treatment, with mean level of DKK1 4.97±2.23 ng/ml pre-treatment and 2.75±1.52 ng/ml post-treatment. These findings agreed with those of Tung et al. [24] and Shen et al. [22] who reported that serum DKK1 levels dropped in HCC patients following surgery. Also, Yamabuki et al., (2007) reported reduced DKK1 serum levels following surgical resection of primary tumors in esophageal squamous cell carcinoma and lung cancer patients [37].

After 1 and 3 months of treatment, there was no statistically significant difference between both groups regarding procedure success, stationary ablation, recurrence, decompensation and survival. Both techniques were successful (83.5% with radiofrequency and 61.1% with ethanol injection).

We found that level of DKK1 was significantly decreased after treatment. DKK1 before treatment was 7.52±1.44 in patients with partial response and 3.88±1.49 in patients with complete response and this suggest that DKK1 may have a prognostic role in predicting treatment response. DKK1 was assessed only one month after treatment where no recurrence is detected during this period with CT. Therefore, we couldn’t emphasize that level of DKK1 elevated again with tumor recurrence.

No studies have been done before to put a cut off value for DKK1 for prediction of treatment response even after surgical resection. In this study, we have a cut off value for DKK1 for prediction of complete response to treatment. This value was ≤5.67 ng/mL (AUC 100%, 95% CI 78.7-100, sensitivity 100% and specificity 100%) and this value need more studies to be confirmed and to prove prognostic role of DKK1 in prediction of treatment response, recurrence and survival.

From this study and its results, serum DKK1 is a secretory protein, it can be easily detected in circulation and it is elevated in HCC cells and not in normal cells. Serum DKK1 could be used to diagnose HCC, especially with inconclusive AFP. Furthermore, serum DKK1 can complement AFP levels to improve the diagnostic accuracy of HCC. DKK1 could predict treatment response and may be a promising prognostic marker for HCC.

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**Conflicts of interest:** None.

**Ethical approval:** The protocol of the study was approved by the ethical committee of Faculty of Medicine, Zagazig University. Informed consents were obtained from all patients.

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